CHARLES P. ALEXANDER

Part I

De ACTA ZOOLÓGICA LILLOANA del Instituto « Miguel Lillo » tomo X, páginas 75-94

T U C U M A N
REPUBLICA ARGENTINA

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New or little-known Tipuloidea (Diptera) from Argentina

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RESUMEN

"Tipuloidea" nuevos o poco conocidos de Argentina (Díptera) I. — En ésta, la primera de una propuesta serie de trabajos sobre los tipúlidos argentinos, el autor da una lista completa de las especies hasta la fecha encontradas dentro de los límites políticos de la República Argentina, con notas sobre algunas especies y describiendo como nuevas Tipula (E.) protrudens, T. (E.) schachovskoyi, Austro-limnophila (A.) neuquenensis y Teucholabis (T.) foersteri.

Our knowledge of the *Tipuloidea*, or crane-flies, of Argentina is still very insufficient and it is certain that many further species remain to be discovered and that great additions will be made to our knowledge of seasonal and geographical distribution. The largest collections and most satisfactory data that we have at present pertain to the vicinity of Buenos Aires; Alta Gracia in Córdoba; the Andean provinces of Catamarca, Jujuy and Tucumán; and the Patagonian territories of Neuquén and Río Negro. Very insufficient collections have been made in Misiones and Tierra del Fuego.

Until 1926, the Diptera, including the *Tipuloidea*, of Patagonia were virtually unknown. In that year Dr. Fred. W. Edwards and Mr. Raymond C. Shannon, with their wives, were in the territory of Río Negro from the end of October to early December, passing into Chile on December 4th. Unusually large collections were made, chiefly in the vicinity of Lago Nahuel Huapí and Lago Correntoso. The *Tipuloidea* of this expedition were discussed by the writer in a volume of the series of reports on the Diptera of Patagonia and South Chile, published by the British Museum (Natural History) ¹.

¹ Alexander, Charles P. Diptera of Patagonia and South Chile. Part. I. — Craneflies, pp. 1-240, pls. 1-12; 1929.

The distinguished entomologist, Dr. Carlos Bruch, became interested in this group of flies about 1920 and provided the writer with abundant material from the vicinity of Buenos Aires and from Alta Gracia, in Córdoba. Also through Bruch, unusually large collections were made available from the Andean provinces of Catamarca, Jujuy and Tucumán, these having been collected by Mr. Vladimir Weiser. At the present time, further important series are available from Mr. Juan Foerster, taken in the vicinity of Buenos Aires and in Misiones; Mr. S. Schachovskoy, in the vicinity of San Martín de los Andes, Neuquén; and from Dr. Petr Wygodzinsky, in Tucumán. The important series of these flies in the collections of the Miguel Lillo Foundation, of the National University of Tucumán, are likewise available and the records of distribution will be included in subsequent papers under this general title,

I wish to express my deep indebtendness to the many persons who have collected specimens of these often neglected insects. Such acknowledgements are made to many friends now deceased, including Brèthes, Bruch, Edwards, Shannon and Weiser, and to other colleagues whose cooperation is gratefully recorded, these including Dr. Eduardo Del Ponte, Sres. Juan Foerster, S. Schachovskoy and Dr. Petr Wygodzinsky. I am further indebted to Dr. Kenneth J. Hayward, Director of the Institute of Entomology of the Miguel Lillo Foundation for his appreciated invitation to prepare this series of papers in which I hope to bring to date our knowledge of the *Tipuloidea* of Argentina.

Among his very numerous important papers on the insects of Argentina, Bruch published two on the $Tipuloidea^2$. The catalogue provides the basic record of this family of flies for Argentina, with a total of 2 species of Tanyderidae, 3 of Trichoceridae, and 222 of Tipulidae reported from Argentina, including a certain number of forms not actually taken in the Republic and still known only from stations in Chile. Since the list was published a certain number of changes in generic and subgeneric assignments have been made so that it appears advisable to prepare a revised list, stressing particularly the distribution of the various species in subgenera, especially in the great genus Tipula which includes

² Bruch, Carlos. Contribución al conocimiento de los Tipúlidos Argentinos (Diptera). — Physis, 17: 3-28, lám. 1-11; 1939.

Catálogo de los Tipúlidos de la República Argentina. — Physis, 17: 29-55, 1939.

approximately one fourth of all known species of these flies in the Republic. In the accompanying list the various forms are numbered and will be so cited in the present report and in subsequent papers of this series. By this means it will be possible to know the total number of species at any time.

Basic list of the Tipuloidea of Argentina (Modified from the Bruch List, Physis, 17:29-55)

It should be noted that this record includes a few species that to this date have not been taken in Argentina (Patagonia) but occur on the Chilean side of Rosales Pass (Llanquihue Province), close to the border, particulary at Casa Pangue on the frontier. It seems certain that such species actually occur on the Argentine side of the Pass and thus they are retained in the list as originally prepared by Bruch. Species that still are known only from Chile are indicated by an asterisk (*).

TANYDERIDAE

- 1. Araucoderus gloriosus (Alexander) *
- 2. Neoderus patagonicus (Alexander)

To this date, the magnificent Tanyderus pictus Phillippi has not been taken in Patagonia but its occurrence is not unexpected.

TRICHOCERIDAE

- 3. Paracladura chilensis Alexander *
- 4. P. edwardsi Alexander
- 5. P. patagonica Alexander

TIPULIDAE Tipulinae

- 6. Valdiviana edwardsina Alexander
- 7. V shannonina Alexander *
- 8. V synempora Alexander
- 9. Holorusia (Holorusia) catamarcensis Alexander
- 10. H. (H.) delpontei Alexander
- 11. H. (H.) fagetorum Alexander
- 12. H. (H.) jujuyensis Alexander
- 13. H. (H.) postnotalis Alexander
- 14. H. (H.) nudicornis (Macquart) (identity uncertain)
- 15. Megistocera longipennis (Macquart)
- 16. Brachypremna australis Alexander
- 17. B. subsimilis Alexander

- 18. Nephrotoma punctifrons (Macquart)
- 19. Tipula (Microtipula) amoenicornis Alexander
- 20. T. (M.) inarmata Alexander
- 21. T. (Eumicrotipula) andalgala Alexander
- 22. T. (E.) andina Brèthes
- 23. T. (E.) antarctica Alexander
- 24. T. (E.) anthonympha Alexander
- 25. T. (E.) austroandina Alexander
- 26. T. (E.) barretoi Alexander
- 27. T. (E.) bigotiana Alexander
- 28. T. (E.) brèthesiana Alexander
- 29. T. (E.) bruchi Alexander
- 30. T. (E.) chilensis Alexander
- 31. T. (E.) chilota Alexander
- 32. T. (E.) crossospila Alexander *
- 33. T. (E.) dimorpha Alexander
- 34. T. (E.) duséni Alexander *
- 35. T. (E.) enderleinana Alexander
- 36. T. (E.) flavoannulata Jacobs
- 37. T. (E.) fuegiensis Alexander
- 38. T. (E.) hylonympha Alexander *
- 39. T. (E.) iquazuensis Alexander
- 40. T. (E.) jacobsiana Alexander
- 41. T. (E.) jaennickeana Alexander *
- 42. T. (E.) lanigera Alexander
- 43. T. (E.) ligulata Alexander *
- 44. T. (E.) magellanica Alexander (includes magellanicala Alexander)
- 45. T. (E.) monilifera Loew (as moniliferoides Alexander)
- 46. T. (E.) nordenskjoldi Alexander
- 47. T. (E.) nothofagetorum Alexander
- 48. T. (E.) nubifera van der Wulp (identity uncertain)
- 49. T. (E.) ona Alexander
- 50. T. (E.) oreonympha Alexander
- 51. T. (E.) parviloba Alexander
- 52. T. (E.) patagonica Alexander
- 53. T. (E.) philippiana Alexander
- 54. T. (E.) pictipennis Walker
- 55. T. (E.) rufirostris Bigot
- 56. T. (E.) tehuelche Alexander
- 57. T. (E.) tristillata Alexander
- 58. T. (E.) varineura Bigot
- 59. T. (E.) wittei Alexander

(Records for *Tipula oblique-fasciata* Macquart are uncertain and may refer to *T. theobromina* Edwards, or to some other species).

60. Pectinotipula argentina (van der Wulp)

Cylindrotominae

61. Stibadocerina chilensis Alexander *

LIMONIINAE Limoniini

- 62. Tonnoiromyia patagonica Alexander
- 63. Limonia (Limonia) catamarcana Alexander
- 64. L. (L.) cerbereana Alexander
- 65. L. (L.) chilensis (Alexander)
- 66. L. (L.) exercita Alexander
- 67. L. (L.) jujuyensis (Alexander)
- 68. L. (L.) nothofagi Alexander
- 69. L. rL.) phatta (Philippi)
- 70. L. (L.) pluvialis pluvialis Alexander
- 70a. L. (L.) pluvialis correntosana Alexander
- 70b. L. (L.) pluvialis fuscolineata Alexander
- 71. L. (L.) polysticta (Philippi)
- 72. L. (L.) praeclara Alexander
- 73. L. (L.) regifica Alexander
- 74. L. (L.) seposita Alexander
- 75. L. (L.) troglophila Alexander
- 76. L. (Rhipidia) bruchiana Alexander
- 77. L. (R.) domestica (Osten Sacken)
- 78. L. (R.) inaequipectinata Alexander
- 79. L. (R.) microsticta Alexander
- 80. L. (R.) subterminalis Alexander
- 81. L. (Dicranomyia) ambigua Alexander
- 82. L. (D.) dissoluta Alexander
- 83. L. (D.) flavofascialis Alexander
- 84. L. (D.) globulicornis Alexander
- 85. L. (D.) infumata (Philippi)
- 86. L. (D.) jörgenseni Alexander
- 87. L. (D.) ohlini Alexander
- 88. L. (D.) omissivena Alexander
- 89. L. (D.) patruelis Alexander
- 90. L. (D.) repentina Alexander
- 91. L. (D.) sanctae-cruzae sanctae-cruzae Alexander
- 91a. L. (D). s. immaculosa Alexander
- 91b. L. (D.) s. vana Alexander
- 92. L. (D.) sibullina Alexander
- 93. L. (D.) subflavida Alexander
- 94. L. (D.) trituberculata ingloria Alexander
- 95. L. (D.) weiseriana Alexander
- 96. L. (Geranomyia) aegualis Alexander
- 97. L. (G.) argentinensis Alexander
- 98. L. (G.) austroandina Alexander
- 99. L. (G.) gaudens Alexander
- 100. L. (G.) platensis Alexander
- 101. L. (G.) serotina Alexander
- 102. L. (G.) subscrotina Alexander
- 103. L. (G.) tibialis Loew)

- 104. L. (G.) valida (Loew)
- 105. L. (Zelandoglochina) angelica Alexander
- 106. L. (Z.) aphanta Alexander
- 107. L. (Z.) bigoti (Alexander)
- 108. L. (Z.) fagetorum Alexander
- 109. L. (Z.) miniata Alexander
- 110. L. (Z.) multiarmata Alexander
- 111. L. (Z.) multinodosa Alexander
- 112. L.)Z.) omissistyla Alexander *
- 113. L. (Z.) pilosipennis Alexander *
- 114. L. (Z.) tehuelche Alexander

Pediciini

- 115. Pedicia (Tricyphona) chilota Alexander
- 116. P. (T.) serrimarga Alexander (as crassipyga Alexander)
- 117. P. (T.) pusitla (Bigot)

Heratomini

- 118. Austrolimnophila (Limnophilella) patagonica Alexander
- 119. A. (Austrolimnophila) bradleyi Alexander
- 120. A. (A.) elnora Alexander
- 121. A. (A.) eutaeniata (Bigot)
- 122. A. (A.) fuscohalterata Alexander *
- 123. A. (A.) hazelae Alexander
- 124. A. (A.) infidelis Alexander
- 125. A. (A.) iris Alexander
- 126. A. (A.) joana Alexander *
- 127. A. (A.) merklei Alexander
- 128. A. (A.) michaelseni Alexander
- 129. A. (A.) microsticta Alexander
- 130. A. (A.) tremula Alexander *
- 131. Epiphragma (Epiphragma) annulicornis Alexander
- 132. E. (E.) solatrix imitans Alexander
- 133. Polymera (Polymera) bruchi Alexander
- 134. P. (P.) obscura Macquart
- 135. P. (P.) unipuncta Alexander
- 136. Mesolimnophila hirsutipes Alexander *
- 137. Limnophila abtrusa Alexander
- 138. L. ctenonycha Alexander
 - (L. flavocauda Bigot status doubtful)
- 139. L. humidicola Alexander
- 140. L. inculta Alexander
- 141. L. melica Alexander
- 142. L. sparsissima Alexander *
- 143. L. subfasciata Alexander
- 144. Shannonomyia antarctica (Walker)
- 145. S. barilochensis Alexander
- 146. S. cacoxena cacoxena Alexander
- 146a. S. cacoxena mendica Alexander

- 147. S. minutipennis Alexander
- 148. Gynoplistia (Paralimnophila) infestiva Alexander *
- 149. G. (P.) irrorata (Philippi) *
- 150. G. (P.) pallitarsis Alexander
- 151. G. (P.) platensis (Alexander)
- 152. G. (Gynoplistia) hylonympha Alexander
- 153. G. (G.) variicalcarata Alexander *
- 154. Hexatoma (Eriocera) andicola (Alexander)
- 155. H. (E.) ogloblini Alexander
- 156. Atarba (Ischnothrix) aetherea (Bigot)
- 157. A. (I.) argentinicola (Alexander)
- 158. A. (I.) fidelis Alexander *
- 159. A. (I.) ignithorax Alexander
- 160. A. (I.) mesocera Alexander
- 161. A. (I.) scutellata Alexander
- 162. A. (I.) tenuissima Alexander

Eriopterini

- 163. Philippiana egregia Alexander
- 164. ? P. pilosipes Alexander *
- 165. Lecteria (Psaronius) abnormis (Alexander)
- 166. Teucholabis (Paratropesa) xystophanes Alexander
- 167. T. (Teucholabis) omissa Alexander
- 168. Gonomyia (Progonomyia) argentinensis Alexander
- 169. G. (P.) illicis Alexander
- 170. G. (P.) maesta Alexander
- 171. G. (P.) pleurolineata Alexander
- 172. G. (P.) saxicola Alexander
- 173. G. (P.) sunchroa Alexander
- 174. G. (P.) thiosema Alexander
- 175. G. (P.) weiseri Alexander
- 176. G. (Lipophleps) adunca Alexander
- 177. G. (L.) bruchi Alexander
- 178. G. (L.) misera Alexander
- 179. G. (Neolipophleps) glabrispina Alexander
- 180. G. (N.) trispinosa Alexander
- 181. G. (Gonomyia) catamarcae Alexander
- 182. Rhabdomastix (Sacandaga) intermedia Alexander
- 183. Cryptolabis (Procryptolabis) argentinensis Alexander
- 184. C. (P.) barilochensis Alexander
- 185. C. (Cryptolabis) atmophora Alexander *
- 186. C. (C.) spatulata Alexander
- 187. Amphineurus (Rhamphoneurus) fuscifusus Alexander *
- 188. A. (R.) glabristylatus Alexander
- 189. A. (R.) nothofagetorum Alexander
- 190. Erioptera (Empeda) basalis (Alexander)
- 191. E. (E.) complicata (Alexander)
- 192. E. (Symplecta) macroptera (Philippi)
- 193. E. (Trimicra) pilipes (Fabricius), var.

- 194. E. (Mesocyphona) fuscivena Alexander
- 195. E. (Erioptera) cladophoroides Alexander
- 196. E. (E.) lunigera Alexander *
- 197. E. (Eriopterella) jaffueli Alexander *
- 198. Tasiocera (Dasymolophilus) brevicornis Alexander *
- 199. Molophilus (Molophilus) appressus Alexander
- 200. M. (M.) araucanus Alexander *
- 201. M. (M.) armatistylus Alexander
- 202. M. (M.) bellicosus Alexander
- 203. M. (M.) bicaudatus Alexander
- 204. M. (M.) breviramus Alexander
- 205. M. (M.) bravispinosus Alexander
- 206. M. (M.) bruchi Alexander
- 207. M. (M.) calceatus Alexander *
- 208. M. (M.) catamarcensis Alexander
- 209. M. (M.) cervus Alexander
- 210. M. (M.) cladocerus Alexander
- 211. M. (M.) clavigerus Alexander
- 212. M. (M.) colossus Alexander
- 213. M. (M.) drepanuchus Alexander
- 214. M. (M.) flavidus Alexander
- 215. M. (M.) honestus Alexander
- 216. M. (M.) inflexibilis Alexander
- 217. M. (M.) laterospinosus Alexander
- 218. M. (M.) perfidus Alexander
- 219. M. (M.) pirioni pirioni Alexander
- 219a. M. (M.) pirioni omissus Alexander
- 220. M. (M.) richardsi Alexander
- 221. M. (M.) rubidithorax Alexander
- 222. M. (M.) stylifer Alexander
- 223. M. (M.) substylifer Alexander
- 224. M. (M.) ternarius Alexander
- 225. M. (M.) tetracanthus Alexander
- 226. M. (M.) tucumanus Alexander

RECORDS AND DESCRIPTIONS

227. Macromastix (Araucomyia) bullocki Alexander

Macromastix (Araucomyia) bullocki Alexander; Ann. ent. Soc. Amer., 24: 622-623; 1931.

The types were from Nahuelbuta, near Angol, Chile, altitude 800-1200 metres, taken among Chilean pines *Arquearia imbricata* Pav., in March. To this date known only from Chile.

2 & &, San Martín de los Andes, Neuquén, January 12-25, 1950 S. Schachovskoy). These specimens differ considerably among

themselves in pattern and indicate an extensive colorational ran-

ge in the species. One is much darker than the other, the entire thorax being almost uniformly brownish black, on the praescutum there being only obscure reddish areas at the anterior ends of the lateral stripes, white the pleura is very extensively darkened, especially on the mesepisternum but excluding the conspicuously whitened dorsopleural area. In the second specimen, the lateral praescutal borders are broadly obscure vellow while on the mesepisternum the vellow color much exceeds the darkened areas which are restricted to the ventral and cephalic parts of the anepisternum and sternopleurite. The venation is shown (Fig. 2). Male hypopygium (Fig. 1) with the caudal margin of the ninth tergite, 9t, with a broad V shaped notch, the setae restricted to the outer two-thirds of the plate, abundant but relatively delicate, directed caudad. Basistyle, b, elongate, with relatively sparse and inconspicuous setae. Outer dististyle, d, long and slender, subcylindrical. Outer dististive longer, terminating in a slender curved subglabrous rod or spine, the lower margin before the rod with numerous short conical spinous points; body of style with abundant long erect to feebly retrorse setae.

(20) Tipula (Microtipula) inarmata Alexander

Tipula inarmata Alexander; Ann. ent. Soc. Amer., 21: 627-628; 1928.

The type was from Puerto Bemberg, Misiones, Argentina, taken Otober 8, 1927, by Raymond and Elnora Shannon. One further 3, likewise from Misiones, February 11, 1950 (Juan Foerster).

(34) Tipula (Eumicrotipula) duséni Alexander

Tivula duséni Alexander: Arkiv för Zoologi, 13 (6): 30-31; 1920.

Described from the Rio Aysén, South Chile, collected by P. Dusén. Other unpublished Chilean records include the following: Angol, April 6, 1935 (D. S. Bullock); Lago Todos los Santos, Llanquihue, mid January 1933 (E. P. Reed); Termas Río Blanca, Cura Cautín, 1050 metres, March 30, 1938 (D. S. Bullock); Pinas de Lonquimai, Nahuelbuta, January 18, 1938 (D. S. Bullock); Valparaiso, April 19, 1932 (E. P. Reed). Although included in the Bruch list there were no definite records for Argentina to the present time.

San Martín de los Andes, Neuquén, January 18-February 2, 1950 (S. Schachovskoy).

(37) Tipula (Eumicrotipula) fuegiensis Alexander

Tipula fuegiensis Alexander; Arkiv för Zoologi, 13 (6): 27-28, pl. 1, figs. 1, 2, 6; 1920. Diptera of Patagonia and South Chile, 1: 64-65, figs. 11, 162, 163, 164; 1929.

The type was from Río Grande, Tierra del Fuego, taken February 1896 by Ohlin. In the Patagonian report above cited I recorded further material from Bariloche and Correntoso, Patagonia, taken by Edwards and Shannon.

San Martín de los Andes, February 2-March 15, 1950 (S. Schachovskoy); Mount Chapelco, 1200 metres, March 12, 1950 (S. Schachovskoy).

(42a) Tipula (Eumicrotipula) lanigera tau Alexander

Tipula lanigera tau Alexander; Bol. Dept. San. Vegetal, 3: 122-124; 1944.

The type was from Temuco, Chile, taken March 17, 1937, by Bullock. Specimens from San Martín de los Andes, Neuquén, and Mount Chapelco, 1200 metres, March 12, 1950 (S. Schackovskoy). Whether this race can be maintained as distinct from the typical subspecies remains in question.

228. Tipula (Eumicrotipula) microspilota Alexander

Tipula microspilota Alexander; Ann. ent. Soc. Amer., 21: 634; 1928.

The type material was from Malleco, Chile, collected in January 1928 by Pirión, and at Lago Panguipulli, Valdivia, Chile, taken February 4, 1924 by Alfredo Faz. Other Chilean records include Aysén, February 1934 (A. Pirión); Lago Todos los Santos, Llanquihue, mid-January 1933 (E. P. Reed); Termas Río Blanca, Cura Cautín, 1050 metres, March 25, 1938 (D. S. Bullock).

San Martín de los Andes, Neuquén, 620 metres, January 1950 (S. Schachovskoy).

229. Tipula (Eumicrotipula) protrudens sp. n.

Belongs to the glaphyroptera group; general coloration of the mesonotum gray, the praescutum with four entire brown stripes, the lateral borders and a capillary median vitta further darkened; antennae short, basal flagellar segments bicolored; wings with a strong brownish yellow tinge, virtually unpatterned; abdominal tergites yellow, trivittate with brown, the sternites with a central

darkened line; subterminal segments dark brown to form a ring, hypopygium brownish yellow; male hypopygium with the caudal margin of tergite produced into truncated lateral lobes and with a small median tooth at the base of a central emargination; appendage of eighth sternite unequally trilobed, the median lobe longest, provided, with strong curved setae.

Male. — Length about 15-16 mm; wing 15-17 mm; antenna about 2.9-3 mm.

Frontal prolongation of head brown; nasus distinct; palpi with the basal segmnt brown, the remainder brownish black. Antennas (male) short, as shown by the measurements; scape brown, heavily pruinose above; pedicel and first flagellar segment yellow, the succeeding two or three segments weakly bicolored, their bases darkened, the stems paler; outer segments uniformly brownish black; flagellar segments very weakly incised; verticils subequal in length to the segments. Head light gray, the vertex with a central brown line and with darkenings behind the eyes; vertical tubercle low.

Pronotum brownish testaceous, with three confluent brown spots-Mesonotal praescutum gray, with four entire brown stripes, the median interspace further divided by a capillary brown central vitta; lateral praescutal borders broadly darkened, the humeral region more reddish brown; scutum light gray, each lobe with two very unequal brown areas, the more cephalic one small; poster or sclerites of notum gray, the mediotergite with a brown central line. Pleura and pleurotergite gray, the meron and dorsopleural membrane yellowed. Halteres with the stem dark brown, very narrowly pale at base, knob darkened. Legs with the coxae gray pruinose, the fore pair somewhat more obscured; trochanters yellow; femora obscure yellow, with a vague to scarcely evident pale brown subterminal ring; remainder of legs brownish yellow, the outer tarsal segments passing into black; claws (male) simple. Wings with a strong brownish yellow tinge, only vaguely patterned with darker; stigma and faint indications of three subcostal areas pale brown; whitish obliterative areas before stigma and across cell 1st M_2 ; veins brown, more yellowish brown in the costal and prearcular fields. Venation: R_s about two and one-half times m-cu; R₁₊₂ entire; petiole of cell M₁ from about one and one-half to approximately twice m.

Basal abdominal tergite darkened, pale laterally; succeeding tergites yellow, trivittate with brown, the lines interrupted by narrow pale posterior borders to the segments; lateral tergal borders pale;

sternites yellow, with a nearly continuous brown central stripe: subterminal seaments dark brown to form a ring: hupopugium brownish yellow. Male hypopygium (Fig. 3) with the ninth tergite. 9t, narrowed basally, widened at and beyond the middle, the posterior horder with two low truncated lobes that are separated by a U-shaped emargination, in the base of which lies a slender tooth. Basistule cut off from the ninth sternite by a distinct suture, the outer end produced into a more narrowed flattened blade. Ventral lobe of basistyle a slender cylindrical lobe, some five or six times as long as thick, provided with long coarse setae, especially on outer third. Mesal lobe of basistyle unusually large and conspicuous, protruding beyond the other elements of the hypopygium as a fleshy subcylindrical lobe, directed caudad, the surface with microscopic pale setulae. Outer dististule, d. unusually slender, the outer half about twice as thick as the base, provided with long setae, Inner dististyle with the beak relatively slender, obtuse, the blackened edge microscopically scabrous: dorsal crest with the surface delicately ribbed or reticulated, the surface with abundant yellow setae that become small and finally lacking near the beak. Eighth sternite, 8s. with a trilobed appendage, the central lobe longest, narrowed outwardly, the apex and central part membranous, pale, with unusually strong curved setac, the lateral margins of the proximal two-thirds of the lobe thicker and darker: lateral lobes broadly triangular. pale. Gonapophysis, g. unusually powerful, shaped about as shown. Holotype, & Mount Chapelco, Neuguén, 1200 metres, March 12,

Holotype, 3, Mount Chapelco, Neuquén, 1200 metres, March 12, 1950 (S. Schachovskoy). Paratopotypes, 4 3.3.

The most similar described regional species include Tipula (Eumicrotipula) microspilota Alexander and T. (E.) triemarginata Alexander, both of which have the wing pattern somewhat as in the present fly, differing very strikingly in all details of structure of the male hypopygium.

230. Tipula (Eumicrotipula) schachovskoyi sp. n.

Belongs to the glaphyroptera group, allied to apterogyne; nearly apterous, the wings, including the male sex, shorter than the halteres; male hypopygium with the tergite large, the posterior border with a broad V-shaped notch; gonapophysis unusually large and powerful, the apex weakly and irregularly bilobed; eighth sternite unarmed.

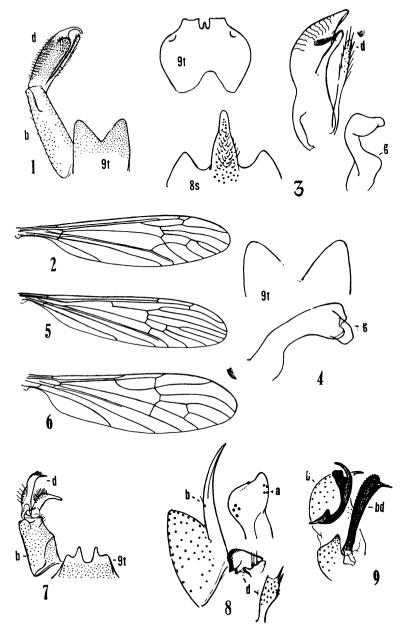


Fig. 1. — Macromastix (Araucomyia) bullocki Alexander; male hypopygium: Fig. 2. — Macromastix (Araucomyia) bullocki Alexander; venation. Fig. 3. — Tipula (Eumicrotipula) protrudens, sp. n.; male hypopygium. Fig. 4. — Tipula (Eumicrotipula) schachovskoyi, sp. n.; male hypopygium. Fig. 5. — Austrolimnophila (Austrolimnophila) neuquenensis, sp. n.; venation. Fig. 6. — Teucholabis (Teucholabis) foersteri, sp. n.; venation. Fig. 7. — Austrolimnophila (Austrolimnophila) neuquenensis, sp. n.; male hypopygium. Fig. 8. — Teucholabis (Teucholabis) foersteri, sp. n.; male hypopygium. Fig. 9. — Molophilus (Molophilus) catamargensis Alexander; male hypopygium.

(Symbols: a, aedeagus; b, basistyle; bd, basal dististyle; d, dististyles; g, gonapophysis; s, sternite; t, tergite).

Male. — Length about 10-11 mm; wing 1.4-1.5 mm; antenna about 4.8-5 mm.

Frontal prolongation of head obscure yellow, heavily gray pruinose; nasus stout; palpi black, the third segment sometimes paler at tip. Antennae (male) relatively long, nearly half the length of the body; scape and pedicel yellow; first flagellar segment bicolored, the proximal third brownish black, the apex obscure brownish yellow; second and third segments less evidently bicolored, the pale color more obscured and decreased in amount; outer flagellar segments uniformly blackened; flagellar segments with moderately developed basal swellings; verticils unusually short, about one-fourth to one-fifth the segments; a conspicuous erect white pubescence. Head brownish gray, clearer gray on front and orbits, obscure yellow on occiput; vertical tubercle very low and simple.

Pronotal scutum light brown, the central line narrowly blackened; surface with scattered black setigerous punctures; scutellum and pretergites yellow. Mesonotum chiefly gray, the praescutal stripes poorly indicated, darker gray; posterior sclerites even more whitened, the mediotergite with a capillary black central line; plurotergite, pale yellow. Pleura gray pruinose, the dorsopleural membrane yellow. Halteres infuscated, especially the knob, the base of the stem narrowly pale. Legs with the coxae and trochanters whitish; remainder of legs brown, the femora a trifle more darkened at tips; tarsi black; claws (male) simple. Wings of male reduced, shorter than the halteres, in cases only about two-thirds as long; venation entirely malformed; wings infuscated, the basal third more yellowed.

Abdominal tergites bright yellow, trivittate with darker; median stripe narrow, black, virtually continuous; lateral stripes much broader but paler, constricted to interrupted at the incisures so the areas appear more or less triangular on the individual segments; the darkened areas become more extensive on the outer segments, the yellow ground correspondingly restricted; extreme tergal borders pruinose; sternites more yellowed, more or less pruinose, with a brown median line, on the outer segments this more expanded behind to form narrow posterior margins instead of a median line; hypopygium pale whitish pruinose. Male hypopygium (Fig. 4) with the ninth tergite, 9t, large, broader than long, the posterior border with a broad and deep V-shaped notch; lateral lobes rather narrowly obtuse at tips which are covered with microscopic setulae. Basistyle

with lobe small, provided with coarse setae. Outer dististyle slender. Inner dististyle much as in dimorpha and subapterogyne, the beak obtuse, densely set with microscopic setae from pale punctures; lower beak and adjacent margin of the upper beak blackened. Gonapohysis, g. unusually strong, the stem stout, the knob weakly and irregularly bilobed. Eighth sternite without an appendage.

Holotype, & Mount Chapelco, near San Martín de los Andes, Neuquén, 1200 metres, March 12, 1950 (S. Schachovskoy). Paratopotypes, 3 & &, one in the collection of Juan Foerster, Buenos Aires.

I take pleasure in naming this interesting fly for the collector, Mr. S. Schaehovskoy, who has discovered several interesting Tipulidae in the vicinity of San Martín de los Andes. This species is readily distinguished from the two most closely allied forms, Tipula (Eumicrotipula) apterogyne Philippi and T. (E.) dimorpha Alexander, by the subapterous condition of the male, and undoubtedly of the still unknown female, and in all details of structure of the male hypopygium. In the lack of an appendage to the eighth sternite, the fly is more like apterogyne, differing in the hypopygial structure, especially the tergite and gonapophysis.

231. Limonia (Zalusa) falklandica (Enderlein)

Zalusa falklandica Enderlein; Zool. Anzeig., 29: 71-72, figs.; 1906.

Originally described from Port Darwin, Falkland Islands, taken on March 3, 1902. Later (*Diptera of Patagonia and South Chile*, 1:85, footnote; 1929) Edwards records further specimens that were taken by the naturalists of the *Challenger* expedition, apparently taken in the Missier Channel, Patagonia, January 1876.

232. Orimarga (Diotrepha) travassosi Alexander

Orimarga (Diotrepha) travassosi Alexander; Rev. de Entomologia (Brasil), 14: 299-300; 1943.

The types were from Juquia, São Paulo, Brazil, altitude 40 metres, taken April 7, 1940, by Lauro Travassos (filho). One male, Misiones, Argentina, February 10, 1950 (Juan Foerster).

(116) Pedicia (Tricyphona) serrimarga Alexander

Pedicia (Tricyphona) serrimarga Alexander; Eol. Dept. San. Vegetal, 3: 129-130, fig. 5; 1944.

Type material from Lago Gutierrez, about six miles south of Bariloche, taken November 11, 1926, by Raymond and Elnora Shan-

non. It had earlier (Diptera of Patagonia and South Chile, 1:110; 1929) been recorded as Pedicia (Tricyphona) crassipyga Alexander, still known only from Chile.

233. Austrolimnophila (Austrolimnophila) neuquenensis, sp. n.

Size medium (wing, male, under 9 mm); mesonotum dark gray, the praescutum with three brownish black stripes, the median one widened on the posterior two-thirds; antennae short black; halteres with stem yellow, knob infuscated; legs brown to brownish black; wings weakly tinted with yellow, restrictedly patterned with brown; male hypopygium with the caudal margin of the tergite irregulary four lobed, the slender inner lobes with smaller lobules on their outer margin; outer dististyle terminating in a powerful black spine.

Male. — Length about 8,5 mm; wing 8,8 mm; antenna about 1,8 mm. Rostrum light ashy gray; palpi black. Antennae (male) relatively short, black, throughout; flagellar segments passing through subcilindrical to more elongate oval, with truncated ends; longest verticils nearly as long as the segments. Head light gray; anterior vertex broad, nearly four times the diameter of the scape.

Pronotum gray pruinose. Mesonotal praescutum dark gray, with three brown to brownish black stripes, the median one much narrower on the anterior third, more expanded behind and here waakly divided by a pale vitta; lateral stripes straight, paler than the central one; scutellum and median region of scutum broadly gray, the scutal lobes almost entirely brownish black; postnotum gray pruinose, the pleurotergite more infuscated. Pleura gray pruinose; dorsopleural membrane more yellowed. Halteres relatively long, stem yellow, knob infuscated. Legs with the coxae yellow, the fore pair and bases of the middle ones sparsely pruinose; femora brown, the bases narrowly more yellowed; tibiae and tarsi dark brown to brownish black. Wings (Fig. 5) weakly tinted with yellow, rstrictedly patterned with brown, this including spots and seams at origin of Rs, forks of Sc and R2+2+4, the latter virtually confluent with a seam over the cord; outer end of cell 1st M2 darkened; stigma small, brown, its proximal half paler; no darkenings at arculus or on the Anal veins; veins brown, restrictedly brightened at wing base. Venation: Sc_1 ending nearly opposite the fork of R_{2+3+4} , Sc_2 near its tip; Rs angulated and short-spurred at origin; R₁₊₂ a trifle longer than R_2 ; R_{2+3+4} about three times the basal section of R_5

petiole of cell M_1 nearly three times m; m-cu just before midlength of cell 1st M_2 .

Abdominal tergites dark brown, the posterior borders of the segments narrowly pale; sternites yellow, the posterior borders narrowly more whitened; eighth and ninth segments blackened. Male hypopygium (Fig. 7) with the caudal margin of the tergite, 9t, irregularly four lobed, there being narrow lobes on either side of a broad U-shaped notch, with small obtuse lateral lobules and their outer margins. Outer distisyle, d, terminating in a powerful blackened spike. Inner disistyle strongly arcuated, its basal third broad, the bent outer part much more narrowed; outer margin of style on basal half with numerous pale erect setae. Phallosome massive, in the unique type more or less folded so that it cannot be well described.

Holotype, &, San Martín de los Andes, Neuquén, January 22, 1950 (S. Schachovskoy).

The most similar species include Austrolimnophila (Austrolimnophila) hazelae Alexander and A. (A.) michaelseni Alexander, which differ conspicuously in the coloration, wing pattern, and in the case of the former species, in the very distinct male hypopygium. The male sex of michaelseni is still unknown.

234. Gynoplistia (Dirhipis) striatipennis Alexander

Gynoplistia (Ctedonia) striatipennis Alexander; Rev. Chil. Hist. nat., 31: 246; 1928.

Described from Panguipulli, Valdivia, Chile, Other specimens from Mancera Island, Valdivia, taken January 1, 1927, by Carlos E. Porter. The subapterous female was first discovered at Cayutue, Lago Todos los Santos, Llanquihue, Chile, on January 13, 1933, by Edwyn P. Reed. Other members of the subgenus have the wings of the female fully developed.

A few males, San Martín de los Andes, Neuquén, January 15, 1950 (S. Schachovskoy).

235. Teucholabis (Teucholabis) foersteri, sp. n.

Allied to tullochi; rostrum yellow; head above chiefly brown; mesonotal praescutum with a polished black discal shield, the humeral and sutural regions yellowed; pleura pale yellow; knob of halteres yellow; posterior tibia (male) with a swollen area before tip; wings

subhyaline, stigma and a seam over the anterior cord dark brown; vein R_5 deflected caudad to the wing tip, cell R_4 thus unusually wide, subequal in marginal extent to R_2 ; male hypopygium with the appendage of the basistyle a long, nearly straight spine, provided with sparse setae.

Male. — Length about 6 mm; wing 5,3 mm.

Rostrum yellow; palpi dark brown. Antennae with the scape and pedicel obscure yellow, the basal flagellar segments brown; flagellum broken beyond the second segment; flagellar segments short-oval to subglobular. Head above brown, paler on front and occiput.

Pronotum obscure vellow. Mesonotal praescutum polished black. the stripes entirely confluent, the lateral ends of the suture, humeral region and median area at the suture yellow: scutal lobes blackened. the median region obscure yellow: scutellum yellow, the parascutella a trifle darker: mediotergite with about the cephalic half orange yellow, the remainder and the pleurotergite black. Pleura pale yellow. with a scarcely differentiated more silvery ventral longitudinal stripe, more expanded on the metapleura, Halteres with stem dark brown, knob yellow; legs with all coxae yellow; posterior trochanters a trifle more darkened; femora yellow basally, the fore pair chiefly blackened: remaining femora with only the tips rather narrowly and indistinctly darkened; fore tibiae and tarsi blackened; middle and hind tibiae yellow, tarsi black: posterior tibiae with a slightly swollen glandular area before tip; posterior basitarsus dilated on nearly the proximal third. Wings (Fig. 6) subhyaline, the prearcular and costal fields even clearer; stigma and a confluent seam on the anterior cord dark brown; cell Sc chiefly darkened; region of the arculus clear; veins dark brown, pale yellow at the wing base. Venation: Sc₁ ending just beyond one-fourth the length of the arcuated Rs. Sc. opposite or just before this origin; R2 and basal section of R5 interstitial; outer half of vein R5 deflected very strongly caudad, ending at the wing tip, cell R4 at margin thus unusually wide, subequal in extent to cell R2; m-cu at or close to fork of M.

Abdominal tergites black; sternites variegated black and yellow, the incisures being broadly of the latter color; hypopygium black. Sternal pocket of fifth segment transverse, the cephalic border with relatively numerous black setae that are directed chiefly caudad, those at the ends somewhat more mesad; no pocket on sixth sternite. Male hypopygium (Fig. 8) with the appendage of the basistyle subbasal in position, appearing as an unusually long and strong

nearly straight spine, narrowed to the acute tip; the fringe of setae present in many species of the genus is here greatly reduced. Outer dististyle, d, broken beyond base on both sides, the preserved part as shown; it is probable that the remainder of the structure is much as in elissa or tullochi. Inner dististyle with the beak bidentate. Aedeagus very obtuse, without apical point, the lower face with eight setae, four in either side (as shown), the upper portion with four more delicate setae

Holotype, &, Misiones, Argentina, February 10, 1950 (Juan E. Foerster): Collector's No 300.

I am very pleased to name this fly for the collector, Mr. Juan E. Foerster, of Buenos Aires. The species is closest to *Teucholabis* (*Teucholabis*) elissa Alexander and T. (T.) tullochi Alexander, as shown by the venation, nature of the abdominal pocket in the male, and in the general structure of the male hypopygium, particulary the inner dististyle and aedeagus. An important distincion is found in the spinous appendage of the basistyle which is here unusually long and nearly glabrous.

(170). Gonomyia (Progonomyia) maesta Alexander

Gonomyia (Gonomyella) maesta Alexander; Ent. News, 32: 73-74; 1921.

The types were from the vicinity of the city of Jujuy, Jujuy, Argentina, taken May 22, 1920, by Weiser. A male, Aguas Blancas, Salta, November 24, 1948 (Petr Wygodzinsky).

(176) Gonomyia (Lipophleps) adunca Alexander

Gonomyia (Leiponeura) adunca Alexander; Ent. News, 32: 294-295; 1921.

The types were from Ampajango. Catamarca, 2300 metres, taken November 30, 1920, and from Punta de Balasto, Catamarca, November 11, 1920, all taken by Weiser. Further records include Bolsón, Catamarca, 2700 metres, March 7, 1924 (Weiser); Amanao, Catamarca, 2500 metres, February 6, 1921 (Weiser); Tucumán, City, October 21, 1948 (Petr Wygodzinsky).

236. Gonomyia (Neolipophleps) acuminata Alexander

Gonomyia (Leiponeura) acuminata Alexander; Proc. Acad. nat. Sci. Ihiladelphia, 77-78, pl. 4, fig. 1; 1921.

Part of the type material was from Famaillá, Tucumán, taken October 12, 1920, by Weiser, the record inadvertently omitted from the basic Bruch list. A male, Aguas Blancas, Salta, November 24, 1948 (Petr Wygodzinsky).

(208) Molophilus (Molophilus) catamarcensis Alexander Molophilus catamarcensis Alexander; Ent. News, 32: 292-293; 1921.

The types were from Punta de Balasto, Catamarca, 2300 metres, taken November 22, 1920, by Weiser. Other Argentine records are from the Quebrada « Volcán », near Agua Amarilla, Catamarca, January 29, 1922 (Weiser) and Tafí del Valle, Tucumán, 2000 meters, September 1949 (Wygodzinsky). The distinctive male hypopygium is shown (Fig. 9).

(212) Molophilus (Molophilus) colossus Alexander

Molophilus colossus Alexander; Diptera of Patagonia and South Chile, 1: 215, figs. 109, 254; 1929.

The type was from the Rio Nirico, east of Bariloche, Río Negro, taken on December 1, 1926, by Fred. W. Edwards. It was found by sweeping small trees, chiefly niri, Nothofagus antarctica Oerst. Other Chilean records in my collection are from Aysén, February 1934 (A. Pirión) and Cayutue, Lago Todos los Santos, February 16, 1938 (Kurt Wolffhügel).

One male, San Martín de los Andes, Neuquén, March 5, 1950 (S. Schachovskoy).

237. Molophilus (Molophilus) shannoninus Alexander

Molophilus (Molophilus) shannoninus Alexander; Journ. N. Y ent. Soc., 55: 182-183; 1947.

The type was from Tucumán, taken in March 1926, by Raymond C. Shannon.

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